In the Specification:

Text added to a paragraph is presented in underlined format, while text to be deleted is presented in strike-through or bracketed format.

Please amend the specification as shown:

Please insert the new Sequence Listing in the specification and delete the earlier Sequence Listing.

Please delete the paragraph on page 2 starting at line 28, and replace with the following amended paragraph:

This invention is used in conjunction with the amplification of a target polynucleotide by **[[by]]** any method. These amplification techniques include PCR, ligase chain reaction (LCR), gap LCR, transcription mediated amplification (TAM), nucleic acid sequence based amplifications (NASBA), and strand displacement amplification (SDA).

Please delete the paragraph on page 3 starting at line 11, and replace with the following amended paragraph:

The term "oligonucleotide" as used herein includes linear oligomers of natural or modified monomers or linkages, including deoxyribonucleosides, ribonucleosides, and the like, capable of specifically binding to a target polynucleotide by way of a regular pattern of monomer-to-monomer interactions, such as Watson-Crick type of base pairing, or the like. Usually monomers are linked by phosphodiester bonds or analogs thereof to form oligonucleotides ranging in size from a few monomeric units, e.g. 3-4, to several tens of monomeric units. Whenever an oligonucleotide is represented by a sequence of letters, such as "ATGCCTG," it will be understood that the nucleotides are in a 5' **to** ~~**fwdarw**~~ 3' order from left

to right and that "A" denotes deoxyadenosine, "C" denotes deoxycytidine, "G" denotes deoxyguanosine, and "T" denotes thymidine, unless otherwise noted. Analogs of phosphodiester linkages include phosphorothioate, phosphorodithioate, phosphoranilidate, phosphoramidate, and the like. Generally, oligonucleotide probes of the invention will have a sufficient number of phosphodiester linkages adjacent to its 5' end so that the 5' **to** ~~**fwdarw**~~ 3' exonuclease activity employed can efficiently degrade the bound probe to separate the reporter and quencher molecules.

Please delete the paragraph on page 6 starting at line 25, and replace with the following amended paragraph:

The shorter **probe [[gene]]** may be prepared as a 5' truncate of the longer **probe [[gene]]**. Or it may be a 3' truncate. A third option is to create the shorter probe by trnacting both the 5' and the 3' end of the longer **probe [[gene]]**. Any one of these three forms of the shorter probe will work. While two or more truncated forms could be used, it is simplest to use just one form, preferably the 5' truncate form.

Please amend Table I as indicated in the attached Table I and insert in the specification on page 8 above Table II.

Please delete the header "CV03.SEQ" and replace with SEQ ID NO: 5 (as shown)
Please delete the first line of sequence (as shown)

Please insert the following paragraph on page 8 between line 3 and Table I (not presently shown but to be inserted as indicated above):

The sequence in Table I is indicated as SEQ ID NO. 5. The HCV oligo in Table II is indicated as SEQ ID NO. 1. The HCVR2 oligo in Table II is indicated as SEQ ID NO. 2. Probe

C1 in Table III is indicated as SEQ ID NO. 3. Probe C2 in Table III is indicated as SEQ ID NO. 4.

Please delete the sentence on page 8 starting just below Table IV, and replace it with the following amended sentence:

Molecular **weights** ~~**weighs**~~ calculated by Gene **Runner** **[[Runer]]** version 3.04 (Hasting Software, Inc.).